Pegylated Arginase I Blunts T Cell Function Through Inhibition of Dendritic Cell Development Paul Kepper, Paul Thevenot, Ph.D, Audrey Lemoine, Paulo Rodriguez, Ph.D Stanley S. Scott Cancer Center, Louisiana State University Health Sciences Center



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Abstract

The development of an immune suppressive microenvironment plays a primary role in the growth of tumors and represents a major obstacle in the success of tumor immunotherapy. The metabolism of the non-essential amino acid L-Arginine (L-Arg) through the enzyme arginase I in myeloid derived suppressor cells (MDSCs) is a fundamental mechanism and prime example of the suppressive immune responses in tumor-bearing hosts. Accordingly, the depletion of L-Arg through a pegylated form of human recombinant arginase I (PEG-Arg I) impaired T cell function and delayed the appearance of graft vs. host disease in mice undergoing mismatched bone marrow transplantation (1). Additional results indicated that PEG-Arg I therapies induced the accumulation of MDSCs, suggesting that PEG-Arg I blocked T cell responses mainly through MDSC promotion (2). However, the specific effect of L-Arg starvation in the maturation and function of myeloid cells remains entirely unknown. In this study, we aimed to determine the effect of PEG-Arg I in the maturation of dendritic cells, the ultimate antigen-presenting cells. We hypothesize that L-Arg deprivation by PEG-Arg I hinders the maturation of dendritic cells, leading instead to the accumulation of their precursors, MDSCs. Therefore, PEG-Arg I-based therapies represent a potential therapy for conditions such as self-reactive immune pathologies or T lymphocyte reactions preceding transplantations. Our results show that the treatment with PEG-Arg I impairs T cell proliferation in vivo through the accumulation of MDSCs. Additional findings also indicated that PEG-Arg I blocked the development of dendritic cells in vitro and significantly inhibited their ability to activate T cells. These results, associated with an increased accumulation of MDSCs, suggest that PEG-Arg I blocked dendritic cell differentiation beyond an MDSC stage. Therefore, the overall results suggest that PEG-Arg I impairs T cell function through an arrest of dendritic cell differentiation. Continuation of our research is expected to have a positive public health impact as the findings could enable the development of new therapies for autoimmune disorders and increase the understanding of the important role of the metabolism of L-Arg in immunity.









Figure 3: PEG-Arg I blocks the ability of dendritic cells to efficiently activate T cells.

B Peg Arg I treatment of dendritic cells delays the ability of dendritic cells to present antigens and activate T cells.



Conclusions

PEG-Arg I inhibits T cell proliferation *in vivo* through the induction of

PEG-Arg I impairs maturation of dendritic cells *in vitro*; resulting in

PEG-Arg I blocks the ability of dendritic cells to efficiently activate T cells.

Future experiments will be done to determine the role of CD11b⁺ GR1⁺ in the decreased function of dendritic cells developed in the presence of peg-Arg I and to characterize the pathways by which peg-Arg I prevents myeloid cell maturation. Another set of experiments will identify the effect of L-Arg deprivation in the maturation of dendritic cells in vivo.

References

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